

PP_IL_D109

MIRNAS BOOST BREAST CANCER AGGRESSIVENESS BY REGULATING S-NITROSOGLUTATHIONE REDUCTASE (GSNOR) LEVELS

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S-nitrosylation is the chemical addition of nitric oxide (NO) to a free thiol (-SH) of a protein's cysteine. Various S-nitrosylated proteins (PSNOs) have been found to impact cancer-related pathways such as proliferation, migration, epithelial-to-mesenchymal transition (EMT), and programmed cell death. NO and PSNOs homeostasis are controlled by several enzymes, including NO-producing enzymes such as NO-synthases (NOSs) and different denitrosylases, with S-nitroso-glutathione reductase (GSNOR) being the most characterized, regulating NO and PSNOs levels. GSNOR represents an important regulator of cellular signaling; specifically, its ablation affects DNA repair, senescence, and global transcription and is often related to different pathological conditions. Interestingly, we found GSNOR to be strongly deregulated in breast cancer (BC), suggesting an important role in this tumor, although both the mechanism of GSNOR downregulation and how this affects BC pathogenicity still need to be fully assessed. Our *in silico* analysis revealed that different micro RNAs that could regulate GSNOR levels are also heavily deregulated in BC and are often associated with cancer progression, suggesting that GSNOR downregulation might represent one of the mechanisms through which these miRNAs exert their pro-tumoral activity. In this work, we aim at defining the processes underlying GSNOR downregulation and its biological relevance to highlight GSNOR as a new target for BC treatments.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.170>

PP_IL_E110

IDENTIFICATION OF NITRATED PROTEINS IN HUMAN THROMBI FROM ISCHEMIC CEREBROVASCULAR EVENTS

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A thrombus is formed from fibrin fibres and platelets and is crucial in the coagulation cascade. Fibrinogen converts into fibrin through enzymatic reactions. Platelets, activated in response to vessel damage, adhere and initiate coagulation forming a stabilized clot. The thrombus contains various proteins including clotting factors, which are crucial for proper formation. Understanding thrombi composition is vital for vascular disease mechanisms and therapeutic interventions. Posttranslational modifications occur during nitroxidative distress, an imbalance in reactive species and antioxidants affecting biomolecules. The addition of a nitro functional group to the ring of tyrosine residues, producing 3-nitrotyrosine, serves as a marker of nitroxidative distress found in cardiovascular and inflammatory diseases. Nitration of fibrinogen, observed in thrombotic diseases like thrombosis and stroke, alters fibrin dynamics. Ischemic stroke, an 85% prevalent type, results from decreased blood supply due to clot or endothelium thickening, triggering nitroxidative distress. Represents the second leading cause of death and a major cause of permanent disability. This work analysed thrombi from ischemic stroke patients, searching for 3-nitrotyrosine-containing proteins as potential biomarkers. Thrombi, obtained post-thrombectomy, underwent immunoprecipitation with an anti-nitrotyrosine antibody, followed by SDS-PAGE and Reverse Phase-Liquid Chromatography-Mass Spectrometry analysis in Dynamic Exclusion Mode analysis. Data were processed using the PEAKEStudio XPro search engine for peptide identification. Thrombi exhibited varied compositions in gel electrophoresis, with different protein patterns in each. Western blot

analysis showed that nitrotyrosine residues and fibrinogen fragments were present. Immunoprecipitation revealed varied molecular weights in thrombi pools, suggesting nitrotyrosine-containing polypeptide capture. RP-LC-MS/MS identified nitrated proteins related to the immune system, oxygen transport, and hemostasis, expanding the range of nitrated proteins in ischemic stroke thrombi. The analysed thrombi showed significant protein content variation. Proteins enriched with an inflammatory profile post-anti-nitrotyrosine treatment suggest potential nitrated biomarkers. These findings enhance ischemic stroke understanding, offering insights into thrombi proteomic diversity and potential applications in biomarker research and clinical practice.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.171>

PP_IL_E111

MODULATION OF OXIDATIVE NEUROMETABOLISM IN ISCHEMIA/ REPERFUSION BY NITRITE

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While nitrite is the typical byproduct of nitric oxide (•NO) autooxidation in biological systems, under some circumstances, it can be advantageous to reduce it “back” to the signaling free radical, providing a different, non-enzymatic route for the synthesis of •NO, certain conditions can favor its reduction “back” to the signaling free radical, offering an alternative non-enzymatic pathway for •NO production. In pathophysiological conditions such as ischemia/reperfusion (I/R), where low oxygen availability limits nitric oxide synthase activity, nitrite reduction to •NO may replace enzymatic production, allowing protective modulation of mitochondrial oxidative metabolism and thus reducing the impact of I/R on brain tissue.

In the current study, we used high-resolution respirometry to evaluate the effects of nitrite in an *in vitro* model I/R using hippocampal slices. We found that reoxygenation was accompanied by an increase in oxygen flux, a phenomenon that has been coined “oxidative burst”. The amplitude of this “oxidative burst” was decreased by nitrite in a concentration-dependent manner. These results support the notion that nitrite mediates a decrease in the hyper-reduction of the electron transport system during ischemia, decreasing the accelerated oxygen consumption that characterizes the reoxygenation phase of I/R that has been associated with an increase in oxidant production. Additionally, a pilot *in vivo* study in which animals received a nitrate-rich diet as a strategy to increase circulating and tissue levels of nitrite also revealed that the “oxidative burst” was decreased in the nitrate-treated animals. These results may provide mechanistic support to the observation of a protective effect of nitrite in situations of brain ischemia.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.172>

PP_IL_E112

DECODING THE IMPLICATIONS OF SUBCELLULAR H₂O₂ DYNAMICS IN A STROKE-ON-A-DISH MODEL USING CEREBRAL MICROVASCULAR ENDOTHELIAL CELLS

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Reactive oxygen species (ROS) play a pivotal role in reperfusion injury following cerebral ischemia, the primary treatment for ischemic stroke. The intricate vascular and cellular mechanisms driving this “reperfusion injury” have proven elusive due to the lack of effective *in vivo* methodologies for measuring ROS. In this study, we employed Hyper7.2 and chemogenetic technologies for multiparametric manipulation of H₂O₂ levels, with the goal of elucidating subcellular redox signals in cultured brain endothelial cells. Our investigation utilized stably Hyper7-expressing human cerebral microvascular endothelial cells (hCMEC/D3) to identify key subcellular sources of H₂O₂ production, encompassing the cytosol,

mitochondria, and nucleus. A comprehensive analysis of acute intracellular H₂O₂ signals in hCMEC/D3, subjected to hypoxia/reoxygenation and glucose deprivation, was conducted. Our results revealed that up to 2 hours of hypoxia exposure at 1 kPa did not induce H₂O₂ increase in any cell compartment within CMEC cells. However, during 3–6 hours of hypoxia exposure, a substantial H₂O₂ increase was specifically identified in the cytosol. Following 24 hours of reoxygenation at 18 kPa, H₂O₂ concentrations reverted to normal basal levels. Notably, elevated H₂O₂ levels were solely detected in the cytosol, not in the mitochondria under any conditions, indicating mitochondria's resilience to hypoxic conditions even after reoxygenation. Examination of glucose variations demonstrated that high glucose (11 mM Glc) and low glucose (Glc deprivation, 5 mM) treatments for 3 hours did not induce H₂O₂ level changes. In contrast, in different cellular compartments, subjected to 3 hours of glucose starvation, exhibited a marked increase in H₂O₂ in the cytosol and nucleus, while the mitochondria remained unaffected. In conclusion, hypoxia does not induce significant H₂O₂ changes yet glucose starvation leads to a marked increase in H₂O₂ levels. This study unveils nuanced dynamics in ROS responses, providing valuable insights for potential therapeutic interventions in stroke-related oxidative damage.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.173>

PP II_E113

REDOX AND LIPIDOMIC ANALYSIS OF HUMAN BRAIN MICROVASCULAR ENDOTHELIAL CELLS (hCMEC/D3) ADAPTED TO PHYSIOLOGICAL OXYGEN LEVELS

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The majority of studies with endothelial and other cell types are conducted in standard culture incubators gassed with 5% CO₂ and room air (18 kPa O₂), well known to induce oxidative stress. Cells *in vivo* experience significantly lower O₂ levels, with brain endothelial cells exposed to ~3–7 kPa. Alterations in lipid metabolism mediated by oxidative stress play a key role in ischemic stroke and also are involved in signal transduction. This study aimed to compare the lipidomic profile and redox phenotype of human brain microvascular endothelial cells line (hCMEC/D3) adapted long-term (5 days) to hyperoxia (18 kPa O₂) or physiological normoxia (5 kPa O₂). Redox stress was significantly increased in hCMEC/D3 cells adapted to 18 kPa compared to 5 kPa O₂, as evidenced by elevated basal intracellular GSH (Fig. 1A) and sulforaphane upregulation of NRF2 regulated NQO1 expression. Lipidomic profiling was performed using Laser Desorption - Rapid Evaporative Ionisation Mass Spectrometry, a novel ambient ionization method developed for rapid, sample-preparation-free analysis of complex biological samples. Samples were fixed, washed to remove media and cells were directly analyzed. Preliminary multivariate statistical analysis revealed 87 spectral features as significantly different between hyperoxia (18 kPa) and physiological normoxia (5 kPa). As shown in Fig. 1C, these features are tentatively annotated as glycerophospholipids, specifically phosphatidylglycerol O-36:1, Phosphatidylinositol 38:4 and 38:3. In ongoing further data analysis we are linking these characteristic shifts with specific metabolic pathways. Our study highlights the critical importance of adapting cells *in vitro* to physiological O₂ levels and provides the first insights into shifts in the complex lipidome of cells exposed to different levels of oxidative stress.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.174>

PP II_E114

PERICYTES ARE IMPLICATED IN NO-REFLOW AFTER CEREBRAL ISCHEMIA

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Incomplete reperfusion of the microvasculature (“no-reflow”) damages salvageable brain tissue after ischemic stroke. Using longitudinal *in vivo* 2-photon single-cell imaging we show 87% of pericytes, perivascular cells regulating microvascular flow, constrict during cerebral ischemia, and remain constricted post-reperfusion. Moreover, we reveal ischemic pericytes are fundamentally implicated in capillary no-reflow by limiting and arresting blood flow within the first 24 hours post-stroke. Despite sustaining acute membrane damage, we observe over half of all cortical pericytes survive ischemia and respond to vasoactive stimuli, upregulate unique transcriptomic profiles and replicate. Finally, we demonstrate delayed recovery of capillary diameter by ischemic pericytes after reperfusion predicts vessel reconstruction in the sub-acute phase of stroke and that pharmacological inhibition of Rho-kinase dilates constricted pericytes thereby increasing cerebral blood flow. Cumulatively, these findings demonstrate surviving cortical pericytes remain both viable and are promising therapeutic targets to counteract no-reflow after ischemic stroke.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.175>

PP II_E115

THE RELATIONSHIP BETWEEN MITOCHONDRIAL OXIDATIVE STRESS AND HUNTINGTON'S DISEASE PROGRESSION

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Huntington's Disease (HD) is an autosomal dominant neurodegenerative disease characterized by the accumulation of mutant Huntingtin (mHtt) protein, which interferes with cellular physiological processes. Although the involvement of complex processes such as excitotoxicity, mitochondrial dysfunction, oxidative stress, transcriptional dysregulation, astrocytic dysfunction, and neuro-inflammation in the development of the disease is known, the underlying molecular mechanisms remain largely elusive. Numerous studies have indicated the significant role of oxidative stress in HD pathogenesis. However, due to the limited temporal resolution and invasive nature of current analysis methods, there is still insufficient knowledge about the dynamic regulation of oxidative stress. In our study, we visualized real-time oxidative stress and mitochondrial dynamics at the single-mitochondrion level in striatal neurons using an *in vitro* HD model with the H₂O₂ biosensor HyPerRed. We showed that mitochondrial trafficking was interrupted, and mitochondrial H₂O₂ increased during the mutant Htt protein aggregation. Time-lapse imaging revealed that the subcellular increase in mitochondrial H₂O₂ levels, dependent on mutant Htt aggregation, was correlated with the disruption of intracellular transport. Moreover, we observed oxidative stress increase at the single organelle level with high spatial and temporal resolution for the first time through genetically encoded biosensors during the formation of Huntington's aggregates. This approach provides a new perspective on understanding the role of oxidative stress in the pathogenesis of neurodegeneration in greater detail and sheds light on potential treatments that can halt or reverse disease progression.

doi: <https://doi.org/10.1016/j.freeradbiomed.2024.04.176>

PP II_E116

NOVEL MIRNA TARGETS AND CHARACTERISATION OF OPTIC NERVE FUNCTION IN A MOUSE MODEL OF HYPOPERFUSION

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Background: Understanding biomarkers of vascular dysfunction as mediators of cognitive decline is essential for dementia research. A promising avenue of