

endothelial cells reveals substantial changes in their proteomics profiles. The significantly differentiated proteins were associated with cellular responses to stress and antioxidative related transcription factors. Assessment of Energy metabolism showed decline of oxidative respiration associated with disrupted mitochondria membrane potential. This data suggests that HyPer7 expression challenges cellular homeostasis. Hence, it is essential to exercise caution when using such tools, and to validate outcomes with complementary techniques to ensure the reliability of the obtained results.

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PP I_D44

ADVANCING SUSTAINABLE BIOFUNCTIONAL TEXTILES: EXPLORING THE ANTI-INFLAMMATORY, ANTIOXIDANT, AND UV-PROTECTION PROPERTIES IN ONION (*ALLIUM CEPA* L.) SKIN EXTRACT

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Skin acts as the body's primary defense against external aggressions. Hence, this study investigates the eco-sustainable production of biofunctional textiles using five cellulosic fabrics: cotton, linen, bamboo, hemp, and nettle. These textiles are produced from aqueous extracts derived from the outer skin of the Dorata di Parma onion variety (*Allium cepa* L.). As concerning dyeing experiments, pre-treatment processes were conducted both with and without the use of alum, tin chloride, or tannic acid. The results of UV-protection factor (UPF) measurements showed a significant enhancement in UPF values for all textiles when treated with tannic acid. Indeed, the values ranged from two times higher (in hemp) to six times higher (in bamboo) compared to the corresponding scoured fabrics. Furthermore, when immersing textiles in artificial sweat to simulate skin contact, cotton, hemp, and nettle showed a higher and uniformly distributed release of phenols, indicating greater biofunctional activity, in comparison to linen and bamboo. It was observed that cell viability of primary human dermal fibroblasts (HDF) and skin keratinocytes (NCTC2455) was not affected from the 'bio-functional sweat'. Phenolic components released by biofunctional textiles in artificial sweat were found to provide significant protection against heat shock in HDF cells. Moreover, in both HDF and NCTC2455 cells, the phenolic components from the textiles reduced ROS levels induced by H₂O₂ stress or LPS-induced inflammation. Cotton and hemp textiles, pre mordanted with tin chloride and tannic acid, exhibited higher effectiveness to reduce oxidative stress in vitro. In conclusion, this study highlights the potential of eco-friendly, coloured cellulosic fabrics containing phenol-rich onion peel as a natural shield against UV radiation. Additionally, these fabrics release antioxidants and anti-inflammatory agents onto the skin through sweat, showcasing their multifaceted benefits.

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REVISITING THE ROLE OF H₂O₂ IN NRF2 ACTIVATION USING AN OXYGEN-INDEPENDENT BIOSENSOR

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NRF2 (Nuclear factor erythroid 2-related factor 2) is a crucial transcription factor that governs the expression of genes involved in oxidative stress response, cellular defense mechanisms, and energy metabolism. Impaired NRF2 function has been implicated in aging and various age-related diseases, including neurodegeneration. Despite its significance, the study of NRF2 in living model systems has been hindered by the lack of suitable tools, leading to discrepancies in the literature. One unresolved question is whether and to what extent H₂O₂, a well-known reactive oxygen species (ROS), activates NRF2. To address this, fluorescent

protein (FP)-based biosensors are promising, yet the oxygen dependency of chromophore formation in prevalent FPs poses challenges under hypoxic conditions, potentially yielding inaccurate results. To overcome this limitation, we developed an oxygen-insensitive Nrf2 biosensor reporter based on miniGFP FP, which incorporates flavin to fluoresce, allowing us to monitor Nrf2 behavior regardless of pericellular oxygen levels. Our preliminary findings indicate that the administration of H₂O₂ whether exogenous (bolus; 1-300 uM, up to 24 hours) or endogenous (using mDAAO enzyme; 0.1-50 mM D-alanine, up to 24 hours) is insufficient to stimulate Nrf2 in human cerebrovascular endothelial cells (D3/hCMEC). More interestingly, treatment with Auranofin (0.1-1 uM, up to 24 hours), a TrxR inhibitor, led to significant increases in Nrf2 levels under normal cell culture oxygen conditions (18% O₂). However, when replicating the experiment in cells adapted to normoxia (5% O₂), no Nrf2 activation was observed. Collectively, our findings underscore two striking facts. Firstly, for H₂O₂ to induce Nrf2 activation, abating the antioxidant machinery is essential. Secondly, in comprehensively studying redox biology phenomena, accounting for oxygen level is indispensable; otherwise, the translational relevance of results to clinical settings may be compromised.

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DEVELOPMENT OF A NON-ANIMAL TESTING METHOD USING GENETICALLY ENGINEERED BIOSENSORS FOR ASSESSING SKIN SENSITIZERS

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Skin sensitizers refer to chemical substances that trigger allergic reactions upon skin contact. The industrialization of these products has led to a significant increase in the prevalence of allergic diseases. These all products need to pass through the assessment process before marketing. Due to substantial limitations in the use of animals for risk assessment, considerable efforts have been dedicated to developing non-animal testing methods such as *in silico* or *in vitro*. Some of these tests, approved by the Organization for Economic Co-operation and Development, OECD, rely on luciferase activation linked to Nrf2 expression. Furthermore, genetically encoded biosensors enable real-time measurement of Nrf2 or even hydrogen peroxide (H₂O₂) levels in direct response to oxidative stress increase in a single cell. This advanced technology also allows for the precise targeting of these sensors to distinct subcellular locales, thereby allowing an understanding of localized cellular responses. Therefore, we aimed to develop a drug screening system utilizing keratinocyte HaCat cell line stably expressing the genetically engineered biosensors that detect the oxidative stress level induced by skin sensitizers using the HyPer7 sensor for H₂O₂ measurement and pTRAF sensor for Nrf2 activity assessment. Our data demonstrated that the expression level of either HyPer7 or pTRAF is significantly increased when we treated these stable cells with a high yet not-toxic dose of sensitizer. Moreover, we explored the impact of oxygen levels on the sensitivity of HaCat cells to skin sensitizers. Remarkably, the concentration lethal under hyperoxia conditions (18 kPa) did not induce cell death under normoxia conditions (5 kPa). These preliminary results demonstrate the potential of our approach as a valuable tool for non-animal testing methods, emphasizing its significance in advancing the field.

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HUMAN 3D FULL SKIN MODELS COUPLED WITH ADVANCED BIOMARKERS ASSESSMENT FOR THE EFFICACY EVALUATION OF TOPICAL REDOX MODULATORS

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