

# Design and characterization of nanocarriers loaded with Levofloxacin for enhanced antimicrobial activity; physicochemical properties, *in vitro* release and oral acute toxicity

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Inorganic and carbon based nanomaterials are widely used against several diseases, such as cancer, autoimmune diseases as well as fungi and bacteria colonization. In this work, Santa Barbara Amorphous mesoporous silica (SBA), Halloysite Nanotubes (HNTs) and Multiwalled Carbon Nanotubes (CNTs) were loaded with fluoroquinolone Levofloxacin (LVF) to be applied as antimicrobial agents. The prepared via adsorption nanocarriers were characterized by Fourier-Transformed Spectroscopy, Scanning Electron Microscopy as well as High Pressure liquid Chromatography. *In vitro* release studies were carried out using Simulated Body Fluid at 37°C and data analyzed by various kinetic models showing slow dissolution over 12-24 hours. Antimicrobial studies showed improved antibacterial activity against *Escherichia coli*, *Enterococcus faecalis*, *Listeria monocytogenes*, *Staphylococcus aureus*, and *Staphylococcus epidermidis* compared to neat nanomaterials. CNTs were found to be the most promising candidates for LVF delivery and they were chosen to be further studied for their acute oral toxicity and histopathological examination using C57/Black mice. Histological examination depicted that drug loading did not affect mice organs morphology as well as hepatocyte degeneration, central vein degeneration and parenchymal necrosis scores. To conclude, the prepared nanomaterials present significant characteristics and can act as antimicrobial drug carriers; CNTs found to be safe candidates when orally fed to mice.

**Keywords:** Carbon nanotubes. Santa barbara amorphous. Halloysite nanotubes. Levofloxacin. Toxicity. Antimicrobial.

## INTRODUCTION

Nanotechnology is still the emerging application field for biomedical and pharmaceutical technology applications. Nanomaterials are claimed to be promising in medical field especially as imaging agents or drug carriers due to their large surface area, improved tissue penetration, drug protection against degradation and control of drug administration (De Jong, Borm, 2008; Sifaka *et al.*, 2016c). With the passage of time, inorganic nanoparticles consist of gold, silver or silica and carbon based nanomaterials like nanotubes, graphenes as well

as fullerenes have been used as advanced drug delivery platforms. Graphene comprised from a mono-layer sp<sup>2</sup>-bonded carbon sheet whereas single-walled carbon nanotubes (SWCNTs) and multi-walled carbon nanotubes (MWCNTs) present a cylindrical tube shape of sp<sup>2</sup> carbon, which roll up single- or multi-layered graphene, respectively. They display several unique physical and chemical characteristics which categorized them as suitable candidates in nanomedicine field (Sifaka *et al.*, 2016b,c). The major concern by the CNTs use is the possible toxicity which is undesirable for biomedical applications. It has been outlined that orally fed CNTs seems to be safe even for a high dose of up to 1000 mg/kg (Kolosnjaj-Tabi *et al.*, 2010). In most cases, CNTs have been used so as to deliver anticancer drugs (Son, Hong, Lee, 2016) or antifungal

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drugs (Pieńko *et al.*, 2017). Although, CNTs present several properties, many researchers studied their possible use for antimicrobial applications. Generally, SWCNTs express stronger antimicrobial property than MWCNTs. The fact that synthesis of SWCNTs is more expensive than MWCNTs, improving antimicrobial property of MWCNTs is mandatory. For this reason, a past study, employed the immobilization via poly(ethylene glycol) of cefalexin antimicrobial drug so as to improve the antimicrobial properties of MWCNTs (Qi *et al.*, 2012).

Recently, nanotubes of halloysite, a low-cost clay mineral that expressed numerous physicochemical properties have been introduced as controlled release drug carrier and other biomedical applications (Hanif *et al.*, 2016; Leporatti, 2017). In particular, halloysite nanotubes are natural green clays presenting a cylindrical structure with high aspect length–diameter ratio. Toxicity of HNTs has not been reported yet and claimed as biocompatible and thus can be used as drug delivery carriers. Antimicrobial carriers of HNTs have been reported in literature. In a recent study, HNTs were loaded with Vancomycin and further encapsulated in Alginate and Gelatin/Alginate gels as wound dressings to sustain the drug release (Kurczewska *et al.*, 2017). Another research team evaluated the enhanced antibacterial activity against *Escherichia coli* incorporating halloysite nanotubes (HNTs) and silver into zinc oxide nanoparticles, since neat HNTs exhibit normal bacteria growth (Shu *et al.*, 2017).

Silica based mesoporous materials like SBA-15 (Santa Barbara Amorphous), MCM (Mobil Composition of Matter) and MCF (Mesoporous Cellular silica Foams) are widely found in the literature considering their large surface volume ratio as well as straight narrow channels which allow active molecules (drugs, proteins etc) to highly be absorbed into their structures (Hudson *et al.*, 2008; Ukmar, Planinšek, 2010). In further, SBA-15 mesoporous silicas are also in a considerable interest due to their great adsorption applications such as removal and determination of many molecules. Additionally, SBA-15 cytotoxicity toward B14 fibroblasts was criticized as low (Ferenc *et al.*, 2015). The current years, such nanostructure mesoporous materials have been investigated as drug carriers especially for anticancer drugs (Rajabi, Fayyaz, Luque, 2017), antifungal or anti-inflammatory and immunosuppressant agents (Ortiz-Bustos *et al.*, 2017). Recently, SBA-15 loaded with Ampicillin, a known antibiotic drug was produced, expressing a burst release of the drug from the matrix (Nairi *et al.*, 2017). Additionally, another team researched SBA-15 loaded with Vancomycin or Rifampin as well as a combination of those. They applied them against *S. aureus* and *S. epidermidis*

indicating a statistically significant reduced biofilm density and fewer viable bacteria (Aguilar-Colomer *et al.*, 2017).

In this work in order to enhance antimicrobial properties of MWCNTs, SBA-15 and HNTs, Levofloxacin (LVF) as model drug was chosen. Fluoroquinolones such as LVF have been extensively employed in genitourinary, respiratory, sinus and skin tract infections management (Siafaka *et al.*, 2016a). Researchers have been proposed several nanocarriers for LVF loading, such as polymeric nanoparticles (Karavolos, Holban, 2016), niosomal carriers, nanofibrous matrices, thermosensitive gels (Alves *et al.*, 2016) etc. However, to the extent of our knowledge non applications of carbon based nanomaterials, or halloysite nanotubes and SBA-15 loaded with Levofloxacin have been found.

Therefore, the aim of this work was the evaluation of these nanomaterials as oral carriers for Levofloxacin. For this purpose, SBA, HNTs and CNTs containing LVF were produced and examined for their *in vitro* release and antibacterial property. CNTs loaded with LVF were concluded as the best candidate was orally administered to C57/Black mice to further evaluate the oral toxicity.

## MATERIAL AND METHODS

### Material

Pluronic P-123 triblock copolymer, tetraethyl orthosilicate (TEOS) used for SBA preparation, multiwalled Carbon Nanotubes (CNTs), Halloysite nanotubes (HNTs) and Levofloxacin (LVF) were purchased by Sigma-Aldrich Chemical Co (Steinheim, Germany). All of the used solvents and reagents were of analytical grade.

### Synthesis route of SBA-15

The SBA-15 mesoporous silica was synthesized in accordance to previous reports (Karakoulia, Triantafyllidis, Lemonidou, 2008). Accordingly, Pluronic P-123 copolymer was solubilised in aqueous hydrochloric acid (HCL) (1M, pH ≤ 1) at 35°C. Afterwards, addition of TEOS followed and the solution was magnetically stirring for 60 minutes. The solid particles obtained by filtration, washed with distilled H<sub>2</sub>O, calcinated in air (550°C, 6 hrs) and mesoporous silica was produced.

### Adsorption of the drug onto the nanocarriers

Similarly to a previous work (Filippousi *et al.*, 2015), 200 mg of LVF was dissolved in 200 ml of

Acetonitrile/Methanol (ACN/MeOH) (10/90). Then, 200 mg of SBA, CNTs and HNTs were added to the drug solution. The suspension was magnetic stirring under nitrogen atmosphere for 24 h. Following the solutions were centrifuged and washed with ethanol in order to remove the unloaded LVF particles from the SBA, HNTs or CNTs surface. In order to identify the adsorption the supernatants were quantified using HPLC system and the % adsorption was calculated, given the known LVF concentration.

### Characterization of the LVF loaded nanomaterials

#### Fourier-Transformed Infrared Spectroscopy

The interactions between carriers and LVF drug were examined via FT-IR spectroscopy study. FTIR-spectrometer (model FTIR-2000, Perkin Elmer, Turkey) was used so as to record the baseline corrected spectra, with an area ranged from 4000 to 400  $\text{cm}^{-1}$ , of neat nanomaterials and loaded with LVF formulations. The resolution set of 2  $\text{cm}^{-1}$  (32 co-added scans).

#### Scanning Electron Microscopy (SEM)

The morphology of the pristine and LVF loaded nanomaterials was performed by using a Zeiss EVO scanning electron microscope (SEM). The accelerating voltage was 20 kV, probe current was 45 nA and counting time was 60 s. All the data were analyzed via Digimizer software in order to assess the mean diameter of nanocarriers.

#### In vitro drug release

*In vitro* release study of the LVF loaded nanocarriers was performed in simulated body fluid at 50 rpm. 100 mg of the formulations were added into 100 mL pH 7.4 Simulated Body Fluid (SBF) solution (Siafaka *et al.*, 2015) with a preserved temperature at  $37 \pm 0.5^\circ\text{C}$  to imitate body temperature. 0.5 mL of sample was removed at a predetermined time interval of 15 minute to 24 h while same volume of fresh medium was replaced. The analysis of the drug content was performed with HPLC. The HPLC system (HP Agilent 1100) consisted of a gradient pump and a UV detector set at 295nm. The used column was a C18, 120 A, 5 $\mu\text{m}$ , 250mm $\times$ 4.6mm and column temperature was 25 $^\circ\text{C}$ . The mobile phase comprised from  $\text{H}_2\text{O}$  (pH=3) / MeOH-(60/6400). The applied injection volume was 10 $\mu\text{L}$ . Calibration curve was constructed by using different concentrations of LVF solution (1, 2, 5 10, 25 and 100 ppm) obtained by dilution with mobile phase of a stock aqueous solution of 200 ppm LVF (Siafaka *et al.*, 2016a).

### Analysis of release data

In order to understand the possible release mechanism of LVF drug, the data from *in vitro* drug release studies was exposed to various mathematical models. The best-fit model was decided based on the highest regression values ( $r^2$ ) for correlation coefficients for formulations. For the evaluation and the determination of the release mechanism, kinetic models like zero order, first order, Higuchi and Hixson Crowell were applied. The analysis was utilized to the release study of drug performed until the 60-80% of drug released. In addition, Korsmeyer-Peppas equation was also used for kinetic analysis. When the mechanism is Case I (Fickian) diffusion, it is very well known that the “n” value is around 0.5 (the exact value depends on the geometry). A value between 0.5 and 0.89 reveals anomalous (non-Fickian) diffusion and a value of 0.89 and above demonstrate a Case II Transport. When the release mechanism is mainly based on Fickian diffusion, a dimension less expression of the Higuchi model can be applied (Siafaka *et al.*, 2016a).

#### Disc diffusion method

*Escherichia coli* ATCC 25922, *Enterococcus faecalis* ATCC 29212, *Listeria monocytogenes* ATCC 7644, *Staphylococcus aureus* ATCC 25923, and *Staphylococcus epidermidis* ATCC 12228 were used to evaluate antibacterial activity of formulations by performing disk diffusion method. All microorganism samples were incubated 24 hours using brain-heart infusion (BHI) broth medium at 37 $^\circ\text{C}$ . Following incubation, bacteria adjusted to 0.5 McFarland turbidity standards ( $10^8\text{cfu/mL}$ ) with McFarland Densitometer (DEN-1, Biosan) in 0.85% sterile saline solution. Microbial suspension was cultured on Mueller Hinton agar (MHA) medium using a swab. The entire surface of the Mueller-Hinton Agar (MHA) petri dish was covered with the required inoculum using a swab, and the petri dish was air dried for 15 min. In a sterile environment, formulation discs were applied 5 $\mu\text{L}$  of every formulation were taken by forceps and placed on surface of the petri dish. 15 minutes later, dishes were incubated at 37 $^\circ\text{C}$  for 24 h. Zone diameters occurred around the discs were measured, recorded and the results were evaluated. The study was conducted as pair-parallel manner (Üstündağ-Okur *et al.*, 2014).

#### Acute oral toxicity study

##### Animals

For this study, CNTs loaded with LVF were chosen

as oral optimal candidate and study for their safety profile. Healthy mice were applied for the acute oral toxicity research. In fact, 21 healthy males and females C57/Black mice with weight of 25 g to 30 g and age of 8 to 10 weeks obtained from the MEDİTAM, Istanbul Medipol University. The experiments were carried out according to Declaration of Helsinki for the procedure of animals in study. The experimental procedure relating to the animals was authorized by Istanbul Medipol University Ethical committee (No.27/12/2017-77) before the study started whereas it was conducted under the internationally accepted principles for laboratory animal use and care.

### Acute toxicity assay

#### *Animal conditions*

Mice located in a room at  $22\pm 2^\circ\text{C}$  with constant humidity. A 12 light/12 dark cycle was used as commonly applied. In further, animals were acclimatized to laboratory conditions for a week prior to experiment start. They had free contact to food and water ad libitum. The mice were transferred to the laboratory 1 h before the test activated. All tests were completed between 09:00 and 12:00 h in normal room light and temperature ( $22\pm 1^\circ\text{C}$ ). Mice were separated to three groups of seven animals each ( $n=7$ ), which were treated with the following:

Group I: Blank CNT group,

Group II: LVF loaded CNT group,

Group III: Control group (Distilled water).

Single doses of 2.8 mg/kg of blank and drug loaded CNT formulations were orally administered to both mice in the treatment groups. The formulations were suspended in a vehicle (distilled water). 0.5 mL of formulation was applied by oral gavage. More seven mice were administered with distilled water and named as the control groups. The mice were observed in detail for any indications of toxicity effect within the first six hours after the treatment period, and daily further for a period of 5 days. All animals were weighed and visual observations for mortality, behavioral pattern, changes in physical appearance, injury, pain and signs of illness were conducted daily during the period.

### Measurement of liver weights

After 5 days period, all the mice were sacrificed and liver was isolated and examined for any lesions. In further, liver was weighed and their features were compared between both treated and control groups.

### Histopathological examination

Liver obtained from the mice were transferred in 10% neutral-buffered formaldehyde solution. After dehydration, the samples were blocked in paraffin. A microtome used to cut 5  $\mu\text{m}$ . sections which further stained with hematoxylin and eosin. Mounted slides were evaluated under a light microscope (Nikon Eclipse Ni research microscope, Tokyo, Japan) and photographed using a digital camera (Microscope Digitale Camera DP70, Tokyo, Japan). The criteria for sections evaluation based on intensity and diffusion of degeneration and pleomorphism in hepatocytes, proliferation in bile duct, cytoplasmic eosinophily, parenchymal necrosis, degeneratin, congestion and thrombosis in central vein, and inflammation in portal space. Lastly, the counted values were summarized by each section whereas the degeneration degree was specified in accordance to a score system. Scoring was done as follow: 0-4 (no, low, moderate, high and very high, respectively as no staning (0), low (1+), moderate (2+), heavy (3+) and very heavy (4+).

### Statistical analysis

The statistical analysis was carried out via GraphPad Prism 7.0 software program. The results were pointed out as mean $\pm$ standard error (SEM). Statistical significance among treated and non-treated groups was analyzed by one-way ANOVA followed by Dunnett's post hoc test. P values $<0.05$  were considered statistically significant. Wherever significance has been confirmed, it is showed by \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ .

## RESULTS AND DISCUSSION

### Physicochemical characterization; FT-IR and SEM studies

FT-IR has been evaluated as one of the most important characterization technique used in pharmaceutical technology field. This is because with FT-IR several interactions between the drug and generally active molecules could be depicted. Consequently, herein the spectra of LVF, blank and drug loaded nanocarriers examined. FT-IR spectrum of LVF (Figure 1) exhibits at 2800-3100  $\text{cm}^{-1}$  characteristic peaks due to aromatic group stretching, at 1730  $\text{cm}^{-1}$  the stretching of carbonyl group whereas at 1294  $\text{cm}^{-1}$  stretching of amines (C-N) is also revealed. The peak at 1085  $\text{cm}^{-1}$  is correlated with C-halogen group presence while at region of 3274-3415

$\text{cm}^{-1}$  hydroxyl groups of  $-\text{COOH}$  are illustrated (Siafaka *et al.*, 2016a).

It has been detailed that pure SBA-15 provides silanol groups on the channel walls, which can form a weak intermolecular hydrogen bond with drugs and further allow storage of high amount of a drug and also could prolong the drug release (Jangra *et al.*, 2016). In case of neat SBA-15, the observed broad band at  $3440 \text{ cm}^{-1}$  is attributed to O-H bond stretching vibration of adsorbed water molecules, while at  $1088 \text{ cm}^{-1}$  can be also clearly identified the corresponding bending vibration mode (Figure 1a). For LVF loaded to SBA formulations, an alteration of the spectrum is specified. More specifically, many of the bands were shifted in lower wavenumbers due to interactions (hydrogen bonding) between the LVF and the mesoporous silica (Nanaki *et al.*, 2017).

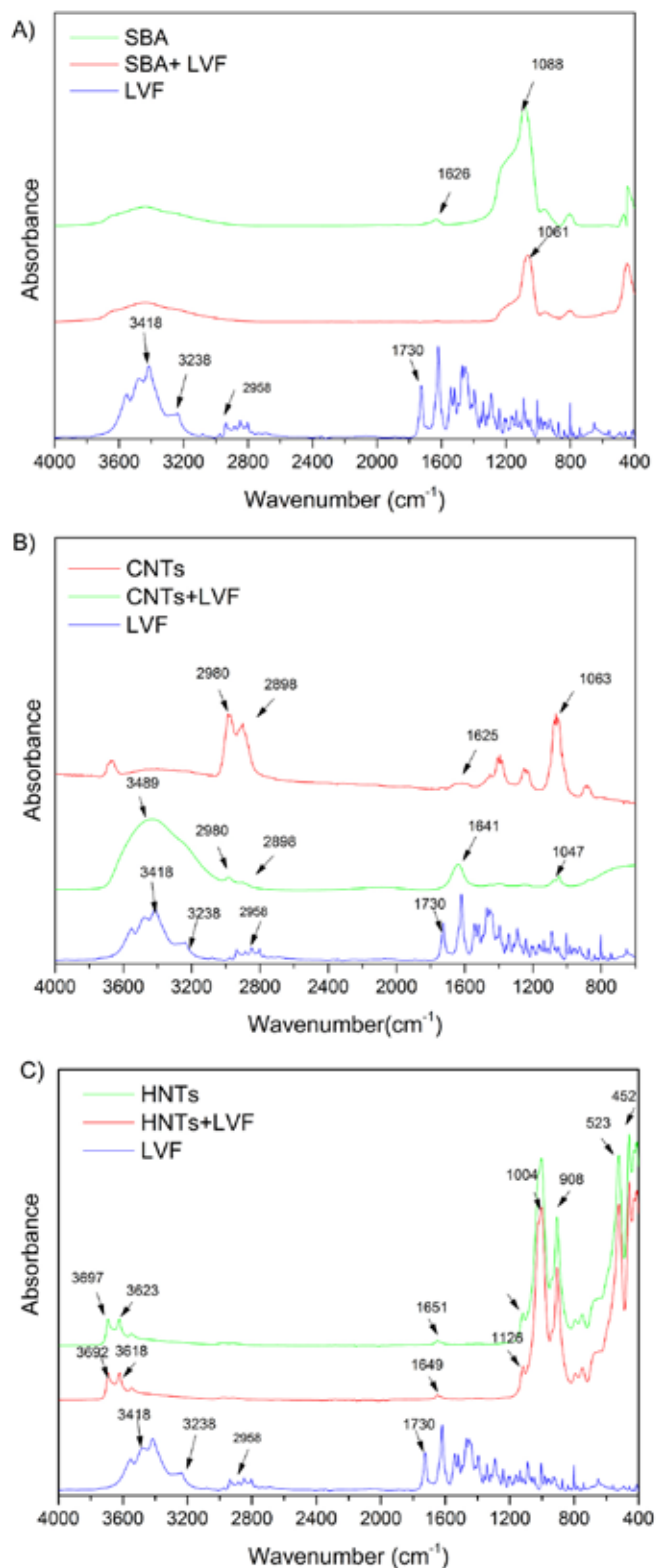
The peaks observed in CNTs spectra at  $1063$ ,  $1625$  and  $3400 \text{ cm}^{-1}$  are correlated with C-O, C=C and O-H bonds (due to water physically adsorbed), respectively (Figure 1a) (Farahani, Behbahani, Javadi, 2016). The band at  $2980$  and  $2898 \text{ cm}^{-1}$  is assigned to asymmetric stretching vibrations of  $\nu(\text{CH}_2)$ . When LVF was absorbed an altered spectrum was seen due to possible interaction between LVF and the nanotubes (Filippousi *et al.*, 2015). Moreover, the broad band at  $3489 \text{ cm}^{-1}$  corresponds to a higher absorbance because of LVF presence.

In case of Halloysite nanotubes the spectra is defined as similar as in the literature reported. For instance, the absorptions bands at  $3697$  and  $3623 \text{ cm}^{-1}$  are due to the stretching vibration of the inner-surface of O-H groups of halloysite, respectively. The vibration at  $1651 \text{ cm}^{-1}$  depicts the interlayer water. The  $1126 \text{ cm}^{-1}$  peak is assigned to Si-O stretching, while the band at  $1004 \text{ cm}^{-1}$  corresponds to Si-O-Si stretching vibration. The band observed at  $523 \text{ cm}^{-1}$  was due to the vibration of Al-O-Si. The vibration of the inner surface hydroxyl group at  $908 \text{ cm}^{-1}$  and Si-O-Si at  $452 \text{ cm}^{-1}$  confirm the existence of corresponding groups. In further, when LVF loaded to HNTs a shifting in lower wavenumbers was also observed indicating interactions between LVF and Si bands (Kurczewska *et al.*, 2017).

SEM analysis was applied in order to morphologically examine the nanocarriers. For pristine CNTs a mean diameter of  $36.8587 \pm 9.5491 \text{ nm}$  was depicted. When LVF was absorbed onto these nanocarriers no morphological alterations was observed whereas drug crystals have not been identified. Given that, LVF drug was not crystallized and thus an increased release rate is found.

SBA-15 mesoporous silicas particles are comprised from tubular mesopores in an ordered hexagonal array (Karakoulia, Triantafyllidis, Lemonidou, 2008). SEM study of the prepared SBA-15 indicated the appearance of

a cluster of particles with mean diameter  $449.5 \pm 107.4 \text{ nm}$ , which are aggregated forming larger clusters of  $4\text{-}5 \mu\text{m}$ .



**FIGURE 1** - FT-IR spectroscopy studies for A) SBA-15 neat and loaded with LVF B) CNTs neat and loaded with LVF and C) HNTs neat and loaded with LVF.

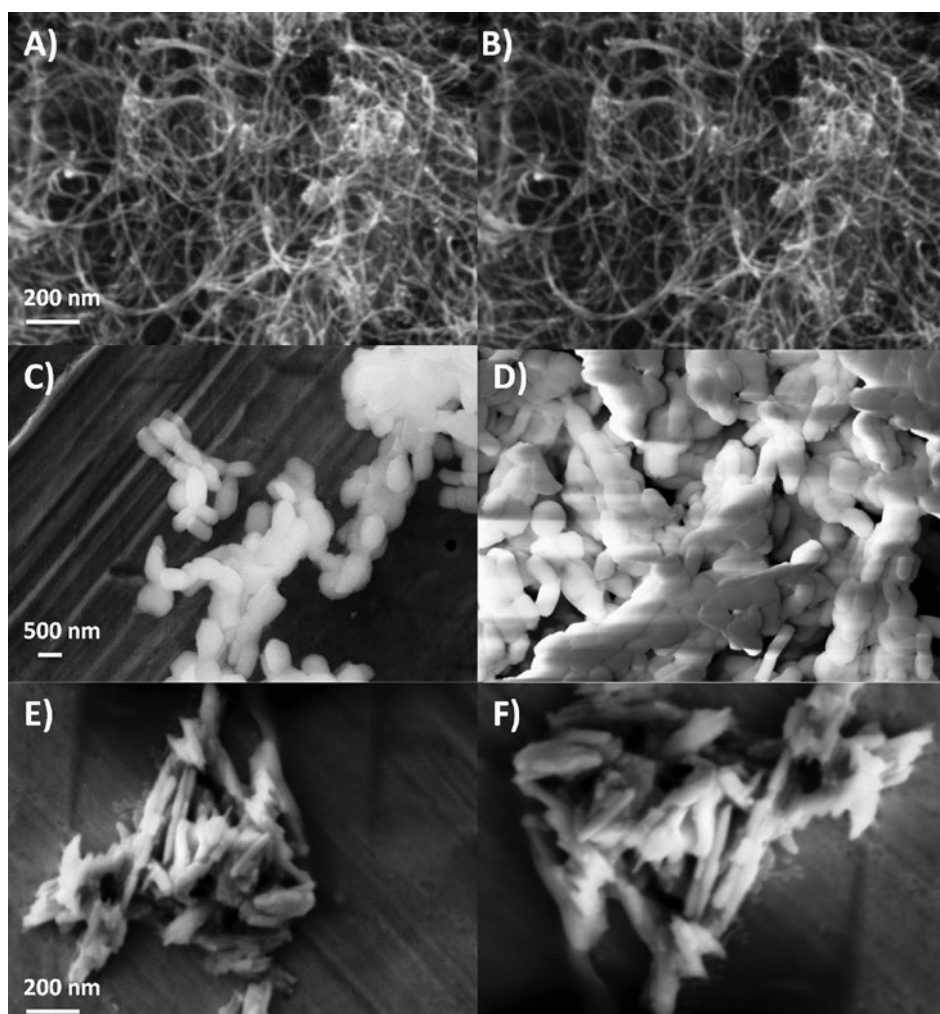
When LVF loaded to the SBA-15 a more compact form was achieved probably due to the drug loading. However, drug molecules were not observed indicating that drug did not crystallize. In case of HNTs a cluster of tubular shaped particles is investigated. As follows, tubes with a mean diameter of  $99.18 \pm 26.31$  nm are observed. Nonetheless, the cluster of these particles presents a diameter of  $2 \mu\text{m}$ . To conclude, when the drug was absorbed the HNTs structure did not affected.

### ***In vitro* release and data analysis**

It has been reported that Levofloxacin is hydrophilic drug. In most cases, hydrophilic drugs express high drug loading however in this work, the drug loading is mostly accelerating by other parameters. When nanomaterials with high surface area ratios as CNTs, HNTs or SBA are used also drug loading is believed to be high. Accordingly,

herein LVF was dissolved using ACN/MeOH solution and nanomaterials were dispersed in the solution in order to obtain high drug loading capacity. The isolation of the materials was achieved via centrifugation and the supernatants were quantified using HPLC in order to find LVF concentration. Table I exhibited the results of the LVF adsorbed nanomaterials. It can be concluded that SBA presents the higher drug loading (23.64%) possibly because of the interactions between LVF and silanol groups, following by CNTs (10.1%) whereas HNTs absorbed only 2.6% of the drug.

Currently, Levofloxacin has been approved as oral, ocular or injectable systems by FDA. LVF is a highly hydrophilic active pharmaceutical ingredient (API) and for this reason was fully dissolved within the first 2 h (data are not shown). However, when the drug is absorbed in the nanocarriers, a slower release rate is observed (Figure 3). It is reported that incorporating a hydrophilic drug onto

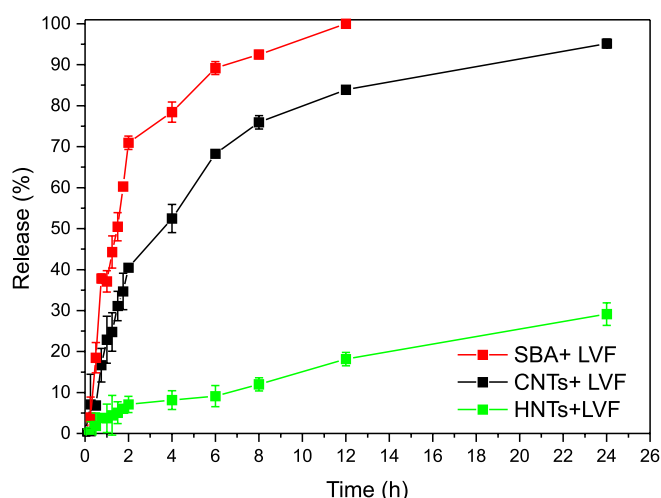


**FIGURE 2** - SEM analysis for A) pure CNTs (original magnification valueX100), B) CNTs loaded with LVF(original magnification valueX100), C) pure SBA-15(original magnification valueX50), D) SBA-15 loaded with LVF(original magnification valueX50), E) pure HNTs (original magnification valueX55), and F) HNTs loaded with LVF(original magnification valueX55).

**TABLE I** - Levofloxacin concentration (%) at filtrates and adsorbed into nanomaterials quantified by HPLC

	CNTs	SBA	HNTs
Levofloxacin theoretical organic solution concentration (mg/mL)	1	1	1
Levofloxacin quantified at filtrate (%)	89.9	76.36	97.4
Levofloxacin adsorbed in nanomaterials (%)	10.1	23.64	2.6

nanomaterials, porous or not, a slower release rate can be achieved. Nonetheless, in almost all cases a burst effect is accomplished (Filippousi *et al.*, 2016; Filippousi *et al.*, 2015).

**FIGURE 3** - *In vitro* release studies of LVF from SBA-15, CNTs and HNTs.

In this study, a low burst phenomenon can be seen for all the nanocarriers, which can be interpreted in regards to the drug deposition on the surface of the nanocarriers. After the initial immediate release, a slower rate is observed. For instance, in case of SBA a more vigorous rate can be observed and 100% of the drug is released in 12 hours. In detail, in 1 hour 37% of LVF was dissolved whereas after 2 hours of dissolution, SBA released almost the 70% of LVF. Finally, 12 hours later complete dissolution is observed. Among others, SBA-15 can control the drug release due to its gate like scaffold

nature in pure and in functionalized form (Jangra *et al.*, 2016). This reduce on dissolution rate is important since an immediate high drug dosage can be toxic to the living organism and thus controlling the rate and administration can be very effective.

Likewise, in case of carbon nanotubes more sustained release is depicted. For instance, after 1 hour almost 23% of LVF was dissolved, in 2 hours dissolution reached at 40% and complete release was achieved after 24 hours. Compared to these nanocarriers, HNTs dissolution profile is completely different. In fact, the first 2 hours of examination only 7% of the drug was released and in 24 hours release rate reached 30%. These observations are rational given that HNTs are used in the literature as controlled release formulations (Cavallaro *et al.*, 2017).

Many past studies examine release mechanism of nanomaterials by various kinetic models. In most of the studies polymeric nanoparticles are evaluated for their mechanism. Diffusion and erosion are the main release mechanisms, found in drug delivery systems. In this work, the data from *in vitro* release studies of formulations were evaluated according to the zero order, first order, Higuchi and Hixson Crowell and Korsmeyer-Peppas kinetic models (Table II). According to the analysis of all mathematical models, it is clearly seen that first order model showed obviously a better fit for all formulations. It has been reported that first-order model is fitted in porous materials loaded with aqueous soluble drugs (Ortiz-Bustos *et al.*, 2017).

Furthermore, to evaluate *in vitro* release kinetics of LVF, the Korsmeyer-Peppas model also examined for LVF loaded formulations. When  $n = 1$ , the release rate is non-dependent of time (zero-order) (case II transport). In

**TABLE II** - Kinetic data of LVF release parameters obtained from formulations.

Models	Zero order			First order			Higuchi			Hixson-Crowell			Korsmeyer-Peppas	
	$r^2$	n	m	$r^2$	n	m	$r^2$	n	m	$r^2$	n	m	$r^2$	n
CNTs+LVF	0.74	24.70	3.796	0.975	4.422	-0.13	0.922	2.34	22.266	0.913	0.364	0.124	0.924	0.79
SBA+LVF	0.74	29.05	9.775	0.954	4.395	-0.32	0.880	2.01	36.380	0.899	0.421	0.320	0.915	1.27
HNTs+LVF	0.97	2.863	1.138	0.982	4.579	-0.01	0.965	-2.59	5.991	0.979	0.042	0.020	0.883	0.76

case of  $n = 0.5$  a fickian diffusion is declared but when  $0.5 < n < 1.0$  both means diffusion and non-fickian transport are involved. Lastly, in case of  $n > 1.0$  super case II transport is apparent 'n' is the slope value of  $\log mt/m\infty$  versus log time curve (Üstündağ-Okur *et al.*, 2015).

In this study, CNTs and HNTs formulations exhibited anomalous (non-Fickian transport) diffusion mechanism with "n" values 0.794 and 0.763, respectively (Table II). The "n" value of SBA formulation has been found to be more than 1. This result indicates that the drug release from the matrix suggests super Case II transport given that these values are characteristic for mass transfer following a non-Fickian model, suggesting that the diffusion of the drug is restricted by the interactions among the drug and the surface silica species (Ortiz-Bustos *et al.*, 2017).

### Antibacterial activity of LVF formulations

Levofloxacin, as it has been reported is a broad spectrum antibacterial agent used to treat numerous bacterial infections such as acute bacterial sinusitis, pneumonia, urinary tract infections, chronic prostatitis, and some types of gastroenteritis (Siafaka *et al.*, 2016a). In addition, it can be used along with other antibiotics to manage tuberculosis, meningitis, or pelvic inflammatory disease. It has been reported that this fluoroquinolone derivative presents high bactericidal activity against *M. Tuberculosis*, *P. aeruginosa*, *S. Aureus*, *Escherichia coli* etc.

Herein, the antimicrobial activity of LVF loaded nanocarriers (Table III) was tested against four opportunistic pathogens commonly associated with oral and pulmonary disease: *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Enterococcus faecalis*, *Listeria monocytogenes*. As it was expected the pristine nanocarriers did not express any bactericidal activity, although it has been claimed that carbon nanotubes could

express antimicrobial activity. However, an inhibition growth was seen from HNTs to E.coli strain (8 mm). In the literature, neat HNTs did not exhibit any bactericidal activity used in its pristine form but when it was decorated with Ag or ZnO showed improved properties (Shu *et al.*, 2017). Statistically significant difference has not been pointed out for antibacterial activity between CNT and SBA against *Staphylococcus epidermidis*, *Escherichia coli*, *Staphylococcus aureus*, *Listeria monocytogenes* ( $p > 0.05$ ). Zone inhibition diameters of LVF solution were determined as 22mm against *Staphylococcus epidermidis* and *Listeria monocytogenes* (Figure 4).

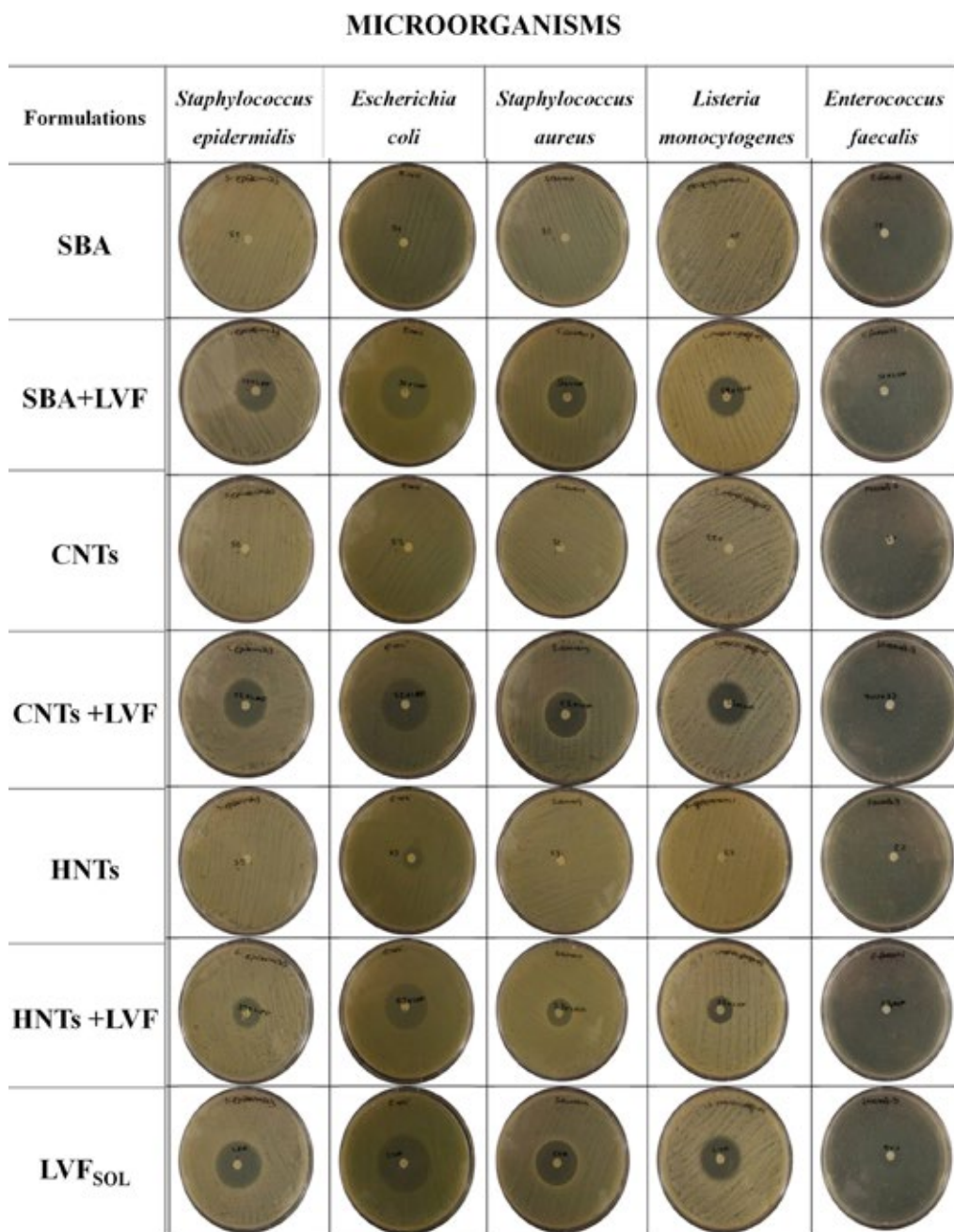
Moreover, a similar inhibition halo (20-32 mm) was observed in case of SBA-15 and MWCNTs loaded with Levofloxacin for all the studied strains revealing that antibiotic activity of such carriers is promising. For example, when SBA loaded LVF was applied against E. coli a higher inhibition halo (32mm) was found compared to CNTs loaded LVF (28mm) which however is not statistically significant. Nonetheless, CNTs loaded with LVF was superior against S. epidermidis, S. aureus and E. faecalis presenting 22, 28, 22 and 32 mm inhibition zone, respectively. SBA loaded LVF and CNTs loaded LVF showed same bactericidal ability in case of *Listeria monocytogenes* (20mm).

It has been revealed, that CNTs immobilized with Cefalexin exhibited also improved bactericidal activity (Qi *et al.*, 2012). Similarly, a past study evaluated SBA-15 loaded with Ag demonstrating increased antimicrobial activity against E. coli due to a synergistic action between extracellular ROS and the toxicity of  $Ag^+$ , which induced intracellular ROS production and subsequent cell death (Wang *et al.*, 2014). In case of HNTs, a slight decreased inhibition halo is seen which is associated with the controlled drug release of the nanocarriers. It can be concluded that compared to free LVF solution a 5%

**TABLE III** - Results of disc diffusion tests (mm)

DIAMETER OF ZONE INHIBITION RING (mm)	Formulations	Microorganisms				
		<i>Staphylococcus epidermidis</i>	<i>Escherichia coli</i>	<i>Staphylococcus aureus</i>	<i>Listeria monocytogenes</i>	<i>Enterococcus faecalis</i>
	SBA	0	0	0	0	0
	SBA+LVF	20	32	18	20	24
	CNTs	0	0	0	0	0
	CNTs +LVF	22	28	20	20	32
	HNTs	0	8	0	0	0
	HNTs +LVF	12	22	12	12	20
	LVF <sub>SOL</sub>	22	30	20	22	30





**FIGURE 4** - Antibacterial activity of the blank and LVF loaded formulations.

reduction in the inhibition zone is seen, which it was expected due to the slower release rate and the difficulty of the drug to be diffused from the carrier (Islan *et al.*, 2016). For the oral toxicity experiment test, CNTs formulations were chosen as the most promising formulations since they also showed better antibacterial activity properties.

**Oral acute toxicity**

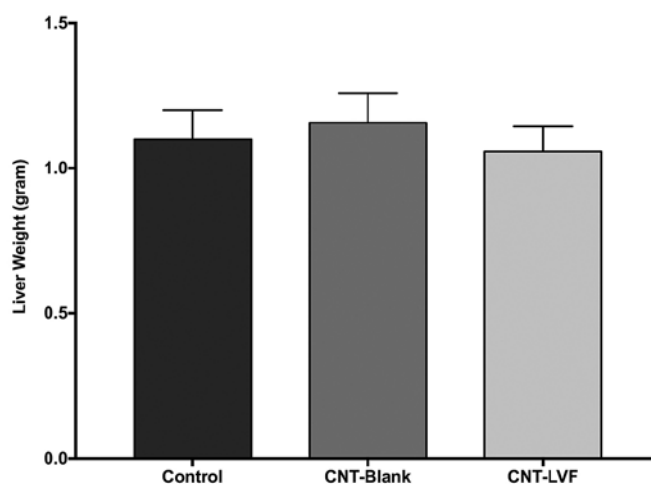
From a scientific aspect, toxicity is the degree to which chemicals or other toxic substances can affect

a living organism. Toxicity can have an impact either on a whole organism (animal, bacteria, and humans) but can also affect a substructure of the organism, such as cells (cytotoxicity) or a bodily organ like the liver (hepatotoxicity) or kidney (nephrotoxicity). Fortunately, living organisms can defend themselves against chemical exposure; for instance, via metabolism, distribution, and excretion various adverse effects can be determined and the risk can be overcome. Nonetheless, exposure in chemicals and drugs, due to renal clearance and detoxification mainly lead to kidney and liver injury (Björnsson, 2016).

As it was referred in this work, CNTs blank and loaded with Levofloxacin were studied for their acute oral toxicity. Throughout literature, CNTs toxicological study reveals conflicting results. Some preliminary *in vitro* toxicological tests suggested that CNTs are toxicologically benign to certain cells, whereas other studies showed that CNTs, mostly raw materials have the potential of being dangerous to many organisms (He *et al.*, 2013). In particular, although CNTs cytotoxicity is not always revealed by the cell culture assay, oxidative stress is the main cause of cytotoxicity (Patlolla, McGinnis, Tchounwou, 2011).

The available safety data indicate that CNTs are of low toxicity via various exposure pathways for biomedical applications. In further, Pulskamp, Diabate and Krug (2007) demonstrated that CNTs did not express acute toxicity in pulmonary epithelial and macrophage cell lines (Pulskamp, Diabate, Krug, 2007). Additionally, an important parameter is the administration dose of CNTs. The most difficult part on CNT toxicokinetics examination is to compare the currently published data due to the differences among the materials. Parameters such as size, dispersibility, functionalization, and metal catalyst contaminants, can lead to discrepancies among toxicity results. Subsequently, detailed chemical and structural characterization of the nanomaterial has to always be given in toxicokinetics assessments of CNT.

In this study, the mice from control and treated groups were orally fed with vehicles, blank and LVF loaded CNTs, respectively. A daily monitoring of the mice was done until 5<sup>th</sup> day in order to observe any toxic signs and mortality. It can be safely reported that toxic symptoms or mortality were not revealed for all animal groups which lived up to 5 days of the experimental period after treated with blank and drug loaded formulations using a single dose level of 2.8 mg/kg body weight. During toxicity evaluation the observation of any clinical symptoms is one of the most significant parameter (Jothy *et al.*, 2011). Mice behavioral patterns were evaluated the first 6 h and 12 h after the administration. It was found that animals in control and CNTs treated groups were normal whereas their behavior, skin, eyes, mucous membrane, sleep, diarrhea, breathing, impairment in food intake and water consumption, postural abnormalities and hair loss have not shown any inconsistencies. Even more, body and liver weight was similar. In detail, in this study liver was not adversely affected throughout the treatment. The liver weight of mice between blank and drug loaded CNTs treated and control groups did not show any statistically significant differences ( $P > 0.05$ ) (Figure 5).



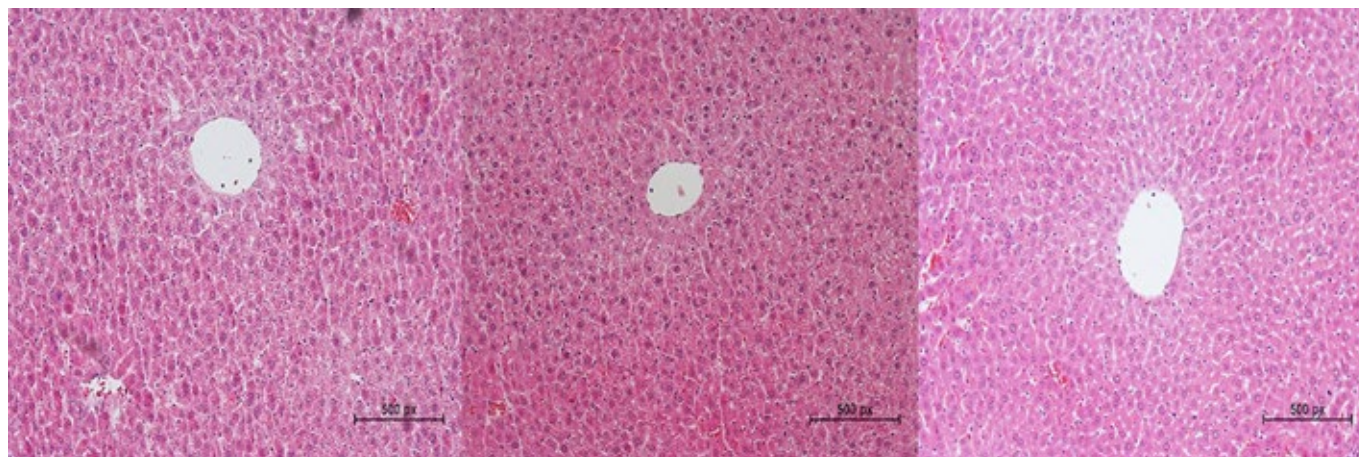
**FIGURE 5** - Comparison of liver weight of mice among groups. Statistically significant as compared to control;  $P < 0.05$  (\*),  $P < 0.01$  (\*\*),  $P < 0.001$  (\*\*\*)

Furthermore, histological examination was performed so as to investigate any alteration in cell structure of the organs. It is well reported that histological study is significant for the evaluation of pathological changes in tissues and organs (OECD/OCDE, 2008). In the present study, histopathological assessment of acute oral ingestion of formulations specified that the formulations did not adversely show impact in the morphology of mice organs. Further, macroscopic investigation of mice organs treated with CNTs did not show any alterations in color compared to control. Finally, autopsy at the end of the experiment did not point out obvious alterations in the liver from both control and treated mice. Figure 5 illustrates the microscopic structures of the organs implying unnoticeable differences between the control and treated mice groups. Likewise, CNTs treated mice organ did not display any changes in cell structure or any unfavorable effects when studied under the light microscope using multiple magnification power. It can be claimed that the structure or coordination of cells in CNTs treated organs are more or less similar compared with the control group organs.

Table IV shows hepatocyte degeneration, cytoplasmic eosinophilia, central vein degeneration and parenchymal necrosis scores of mice livers after treatment 5 days with formulations and control group. All hepatocyte degeneration, cytoplasmic eosinophilia, central vein degeneration and parenchymal necrosis scores were found less than 1. Hepatocyte degeneration and parenchymal necrosis of blank and drug loaded CNTs were found as 0.571. Blank and LVF loaded CNTs scores present same scores except cytoplasmic eosinophilia scores. It can be pointed out that the drug loading have not shown any effect

**TABLE IV** - Hepatocyte degeneration, cytoplasmic eosinophilia, central vein degeneration and parenchymal necrosis scores of the formulations after treatment 5 days

Groups	Control	CNT-Blank	LVF loaded CNT
Hepatocyte degeneration	0	0.571±0.202	0.571±0.202
Cytoplasmic eosinophilia	0	0.714±0.184	0.857±0.261
Central vein degeneration	0	0.429±0.202	0.429±0.202
Parenchymal necrosis	0	0.571±0.202	0.571±0.202

**FIGURE 6** - Histological examination of liver A) blank CNTs B) LVF loaded CNTs C) Distilled water (control group) 20 x (magnification value).

on hepatocyte degeneration, central vein degeneration and parenchymal necrosis scores.

## CONCLUSION

This study aims to check suitability of CNTs, HNTs and SBA carriers as oral drug delivery platforms. The prepared LVF loaded formulations showed improved antibacterial property against several strains. FT-IR spectroscopy revealed interactions between the nanocarriers and the LVF drug while SEM images indicate slight alteration of their morphology after drug loading in case of SBA carriers. From the preliminary studies, LVF loaded CNTs were selected as best formulation. Histological examination of CNTs loaded with LVF showed that drug loading did not affect the hepatocyte degeneration, central vein degeneration and parenchymal necrosis scores. In further, histopathological examination of acute oral ingestion of formulations illustrated that the formulations did not reveal adversely effects on the morphology of mice organs. To conclude, the above nanomaterials showed interesting characteristics and could be an alternative option as antimicrobial drug carriers in biomedical applications.

## CONFLICT OF INTEREST

No declaration.

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